

New saponin from the stem bark of *Samanea Saman*

Nidhi Srivastava

Department of Chemistry, P.P.N. College, Kanpur-208001

E-mail: nidhi.srivastava2006@gmail.com

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Abstract: From the stem bark of *Samanea saman* new triterpenoid glycoside have been isolated and characterized as $3\beta, 22\beta$ – dihydroxyolean -12-ene-24-*O*- β -D-galactopyranoside **1** by spectral data and chemical studies.

(Keywords: *Samanea saman*, new triterpenoid glycoside)

Introduction

Samanea saman (Mimosaceae) commonly called “Raintree” is a useful medicinal plant distributed from Yucatan Peninsula, Guatemala to Peru, Bolivia, Brazil, throughout the West Indies, in old world tropics and also in Southern Florida¹. It is used as an alcoholic source². It is used for cold, diarrhea, headache, intestine ailments, stomachach³, stomach cancer⁴, sore throat⁵. Earlier alkaloids, lupeol, lupeone, octacosanolic acid, hexacosanol, flavonoids, kaempferol⁶ have been isolated from the different parts of *Samanea saman*.

In continuation of our research on the chemical investigation of medicinal plants, we report herein isolation and structure elucidation of new triterpenoid glycosides from *Samanea saman*.

The water insoluble portion of the hot ethanol extract of the air-dried crushed and defatted stem barks of *S. saman* on column chromatography yield compound **1**.

Experimental section :

The stem bark of *Samanea saman* was collected in April, 1999 from the Botany

Department, University of Allahabad, Allahabad, U.P., India. M.ps measured in open capillary tube and are corrected. ¹HNMR and ¹³CNMR spectra were recorded on a JEOLUNK-A 500 spectrometer in CDCl₃ using TMS as an internal standard at 300 MHz and 100 MHz, respectively. IR spectra were run in KBr pellets. Mass spectra were recorded on JEOL SX 102/DA-6000 mass spectrophotometer.

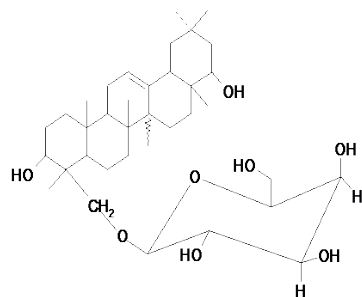
The air-dried and finely crushed stem bark (5kg) of *Samanea saman* was repeatedly extracted with boiling EtOH (4x10l), concentrated under reduce pressure in a rotatory evaporator and poured into an excess of ice-cold distilled water with constant stirring to give reddish brown aqueous solution (fraction-I) and light brown residue (fraction-II) were obtained. The water insoluble portion was extracted successively with different solvents of increasing polarity over a sintered Column. Elution with solvent system C₆H₆: methanol (7:3, v/v) followed by runing in preparative TLC using benzene :chloroform (8:2, v/v) marked with the help of UV lamp gave compounds **1** (0.55g).

Compound **1**, mp. 166°C, yield 550 mg, homogenous on TLC, R_f 0.48 solvent C₆H₆:CHCl₃ (8:2, v/v); Anal. Found: C, 71.90; H, 9.83; Calcd. for C₃₆H₅₉O₈ : C, 71.95; H, 9.89%. IR (KBr) 3450, 2923, 1630, 1100, 1050. ¹HNMR (CDCl₃, 300 MHz) : δ 0.85 (s, 3H, H-25), 0.95 (s, 3H, H-26), 0.99 (s, 3H, H-29), 1.03 (s, 3H, H-30), 1.17 (s, 3H, H-27), 1.31 (s, 3H, H-28), 1.35 (s, 3H, H-23), 2.59 (brs, 1H, J=13Hz), 3.83 (dd, 1H, J=9.6 Hz, β -OH at C-3), 5.25 (t, 1H, J=3.3 Hz), 4.35 and 4.52 (d, 1H, J=12.1 Hz for -CH₂OH group proton), 4.42 (d, 1H, J=7 Hz for

β linkage) and 3.52-3.88 (m, 5H, sugar protons); 204, 201, 175, 119; ^{13}C -NMR data are given in MS : m/z 617[M⁺] 487, 438, 391, 234, 224, 219, 216, **Table-I**.

Table-I
 ^{13}C -NMR (δ) values of glycoside 1 and its aglycone 1a

Chemical Assigned	Glycoside Chemical Shift (ppm)	Aglycone Chemical shift (ppm)
C-1	38.6 (t)	39.0(t)
C-2	27.5(t)	28.4(t)
C-3	80.0 (d)	79.8(d)
C-4	43.0 (s)	43.0(s)
C-5	57.0(d)	56.3(d)
C-6	19.2(t)	19.4(t)
C-7	33.7(t)	33.5(t)
C-8	40.1(s)	39.9(s)
C-9	48.6(d)	48.1(d)
C-10	36.8(s)	37.0(s)
C-11	24.6(t)	24.0(t)
C-12	123.0(d)	122.4(d)
C-13	145.0(s)	144.9(s)
C-14	42.0(s)	42.3(s)
C-15	26.0 (t)	26.4(t)
C-16	29.4 (t)	29.0(t)
C-17	37.5 (s)	38.4 (s)
C-18	44.5(d)	44.8(d)
C-19	41.9 (t)	41.5 (t)
C-20	30.5(s)	30.0(s)
C-21	37.0(t)	37.3(t)
C-22	75.6(d)	75.6(d)
C-23	23.1(q)	23.5(q)
C-24	74.5 (t)	64.5 (t)
C-25	16.2(q)	16.3(q)
C-26	17.5(q)	17.1(q)
C-27	26.8(q)	25.5(q)
C-28	21.1(q)	21.1(q)
C-29	32.5(q)	32.0(q)
C-30	24.6(q)	24.4(q)
C-1'	106.8(q)	
C-2'	74.5 (q)	
C-3'	75.2 (d)	
C-4'	71.1 (d)	
C-5'	76.6 (d)	
C-6'	64.6(t)	



3 β , 22 β – dihydroxyolean -12-ene-24-O- β -D-galactopyranoside **1**

Results and Discussion

Compound **1** ($C_{36}H_{59}O_8$), a glycoside on acid hydrolysis gave D-xylose and an aglycone **1a**. The aglycone gave colour reactions⁷⁻¹⁰ of unsaturated pentacyclic triterpenoid. It formed triacetate on acetylation showing the presence of three hydroxyl group. The IR spectrum of **1a** showed absorption for hydroxyl (3450 cm^{-1}), trisubstituted double bond (1630 cm^{-1}), primary and secondary alcoholic group ($1250, 1100, 1050\text{ cm}^{-1}$) and C-methyl ($2900, 1385$ and 1330 cm^{-1}) groups. The $^1\text{H-NMR}$ spectrum of **1a** showed signals for seven tertiary methyl groups at ($\delta 0.85, 0.95, 0.99, 1.03, 1.17, 1.31$ and 1.35 (each 3H, s) and a vinyl proton at 5.25 (1H, t, $J=3.3\text{ Hz}$) all of which suggested **1a** to be an olean-12-ene derivative.

The mass spectrum of **1a** revealed important peaks at m/z 234, 224, 210, 216, 204, 201 and 175 typical of retro-Diels-Alder fragmentation

of ring-C of olean-12-ene derivative containing a hydroxyl group in ring A or B¹¹. Compound **1a** also gave positive Zimmermann test¹² suggesting the C-3 position for this hydroxyl group which was also biogenetically favoured. The aglycone showed signal at $\delta 3.83$ (1H, dd, $J=9.6\text{ Hz}$) with higher value of coupling constant confirming the β -orientation for hydroxyl group at C-3 (α or axial H)¹³. The ion peak at m/z 216 [$234 - \text{H}_2\text{O}$] due to ready loss of water molecule showed the position of -OH group at C-22 which was also supported by the $^{13}\text{C-NMR}$ signal at $\delta 75.6$ (d) for C-22. The $^1\text{H-NMR}$ signals at $\delta 4.35$ and 4.52 (each 1H, d, $J=12.1\text{ Hz}$) were attributed to the methylene protons of hydroxymethyl group (AB system)¹⁴. Signal at $\delta 64.5$ in $^{13}\text{C-NMR}$ spectrum of **1a** confirmed the presence of CH_2OH group at C-24. Thus, aglycone **1a** was identified as 3 β , 22 β , 24-trihydroxyolean-12-ene.

The $^1\text{H-NMR}$ spectrum of **1** showed a signal for an anomeric proton at $\delta 4.42$ (1H, d, $J=7.0\text{ Hz}$, H-1, galactose) and sugar protons at $\delta 3.52\text{--}3.88$ (br, galactosyl protons) which was consistent with configuration of D-galactose.¹⁵ The β -type of glycosidic linkage was also confirmed by hydrolysis of **1** with the enzyme emulsin. The position of attachment of D-galactose at C-24 hydroxyl group was confirmed by down field signal at $\delta 74.5$ (t) in the $^{13}\text{C-NMR}$ of **1** for C-24. Thus the compound **1** was characterized as 3 β , 22 β -dihydroxyolean-12-ene-24-O- β -D-galactopyranoside.

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